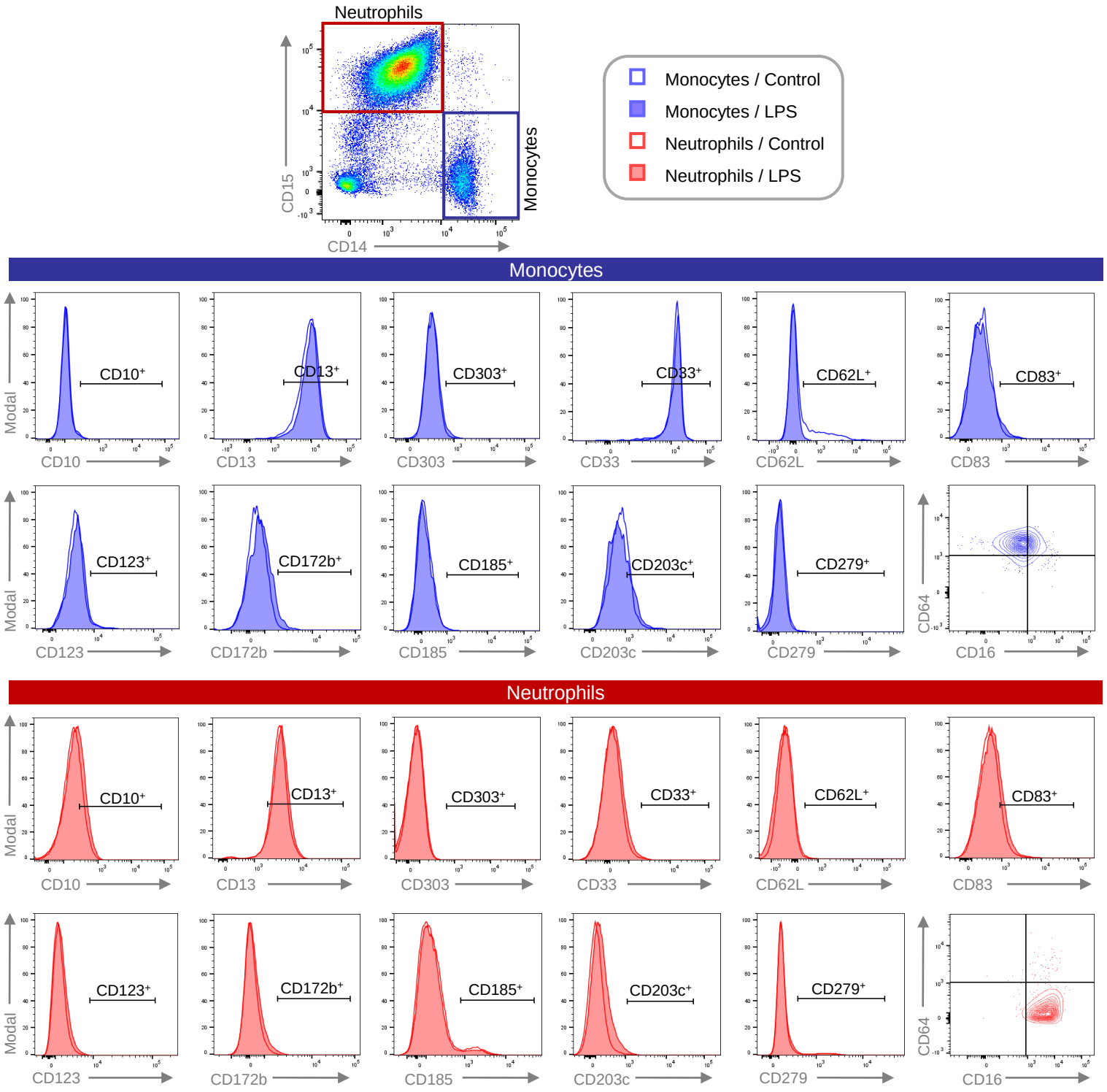
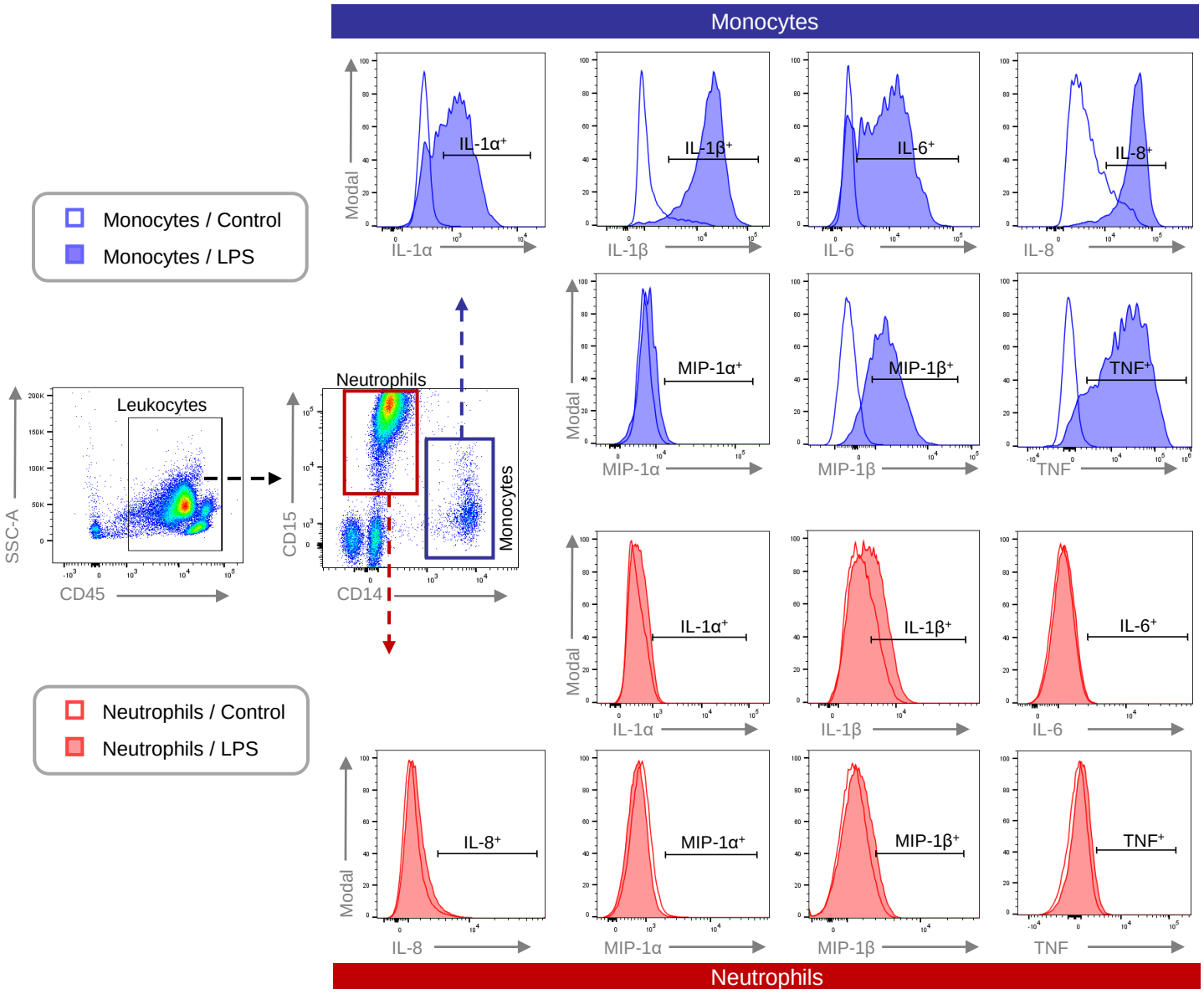
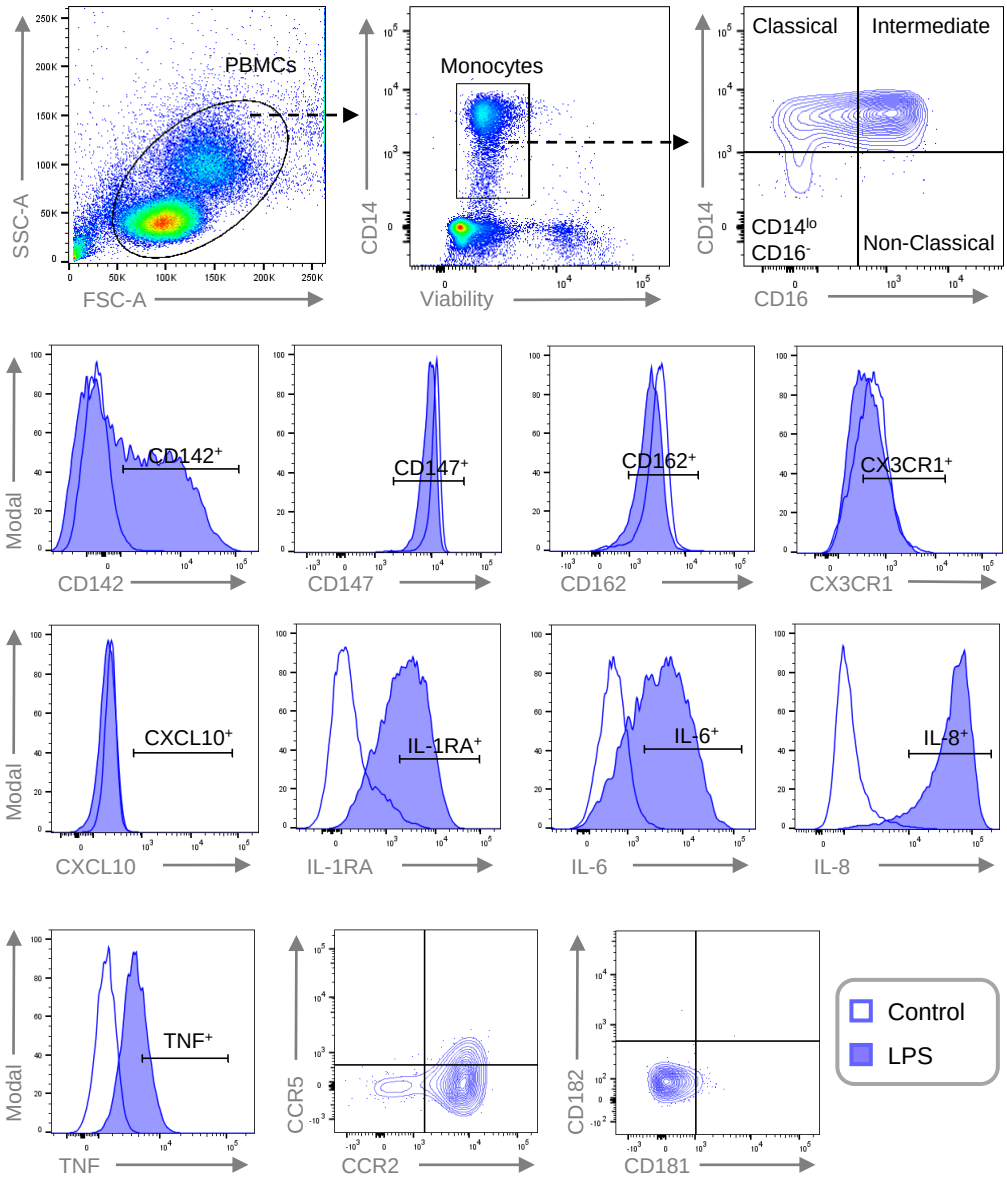


Supplemental Fig. 1 Flow cytometry gating strategy for phenotyping monocytes and neutrophils upon acute LPS stimulation. Monocytes and neutrophils isolated from pregnant (n = 18) and non-pregnant (n = 17) women were gated as $CD14^+$ and $CD15^+$ cells, respectively (described in Fig. 2b&c). Monocytes were gated for immune markers after 4 h of LPS stimulation (filled blue histograms) or control (open blue histograms). Neutrophils were gated for the same immune markers after 4 h of LPS stimulation (filled red histograms) or control (open red histograms).



Supplementary Fig. 2 Flow cytometry gating strategy for cytokine expression by monocytes and neutrophils upon acute LPS stimulation. Leukocytes isolated from pregnant (n = 18) and non-pregnant (n = 17) women were gated for $CD14^+$ monocytes and $CD15^+$ neutrophils (described in Fig. 2d-i). Monocytes were then gated for cytokines and chemokines after 4 h of LPS stimulation (blue filled histograms) or control (blue open histograms). Neutrophils were gated for the same cytokines and chemokines after 4 h of LPS stimulation (red filled histograms) or control (red open histograms).



Supplementary Fig. 3 Flow cytometry gating strategy for phenotyping of monocyte subsets upon chronic LPS stimulation. Peripheral blood mononuclear cells (PBMCs) isolated from pregnant (n = 20) and non-pregnant (n = 20) women were gated for monocytes ($CD14^+$ cells, described in Fig. 3). The expression of CD16 and CD14 was used to gate monocyte subsets as follows: classical ($CD14^{hi}CD16^-$ cells); intermediate ($CD14^{hi}CD16^+$ cells); non-classical ($CD14^{lo}CD16^+$ cells); and $CD14^{lo}CD16^-$ monocytes. All four subsets as well as the total monocyte population were gated for specific phenotypes after 24 h of LPS stimulation (blue filled histograms) or control (blue open histograms).